

# Legacy Arsenic Contamination and its Effects on Gene Expression of the Heat Shock Protein in Sunfish

Jaskiran Dulai, Madeline Hiller, Dr. Alison Gardell

## INTRODUCTION

This study aimed to look at the effects of arsenic from three different lakes with varying levels of arsenic contamination and the effects it can have on the *hsp-70* gene expression of the liver on Bluegill (*Lepomis macrochirus*) and Pumpkinseed Sunfish (*Lepomis gibbosus*). Our proposed hypothesis was:

- The lake with the highest contamination (Lake Killarney) was expected to have a greater *hsp-70* expression than fish with no arsenic exposure from Pine Lake.

Arsenic contamination has been a significant problem in the South Puget Sound area for a long time, a major way arsenic has contaminated the soil and water in Tacoma is from the Ruston copper smelter (Arsenic, 2022).

We chose to focus on the liver in study because it is a major site for detoxification in the body.

## METHODS AND MATERIALS

Samples were taken from 3 lakes- Lake Killarney, Steel Lake, and Pine Lake.

Bluegill and Pumpkinseed Sunfish were collected from the lakes. Fish were euthanized and target tissues were collected (liver)

Tissue extracted using Trizol reagent and homogenized with liquid nitrogen

Purity and concentration was taken- threshold of purity concentrations was >1.7

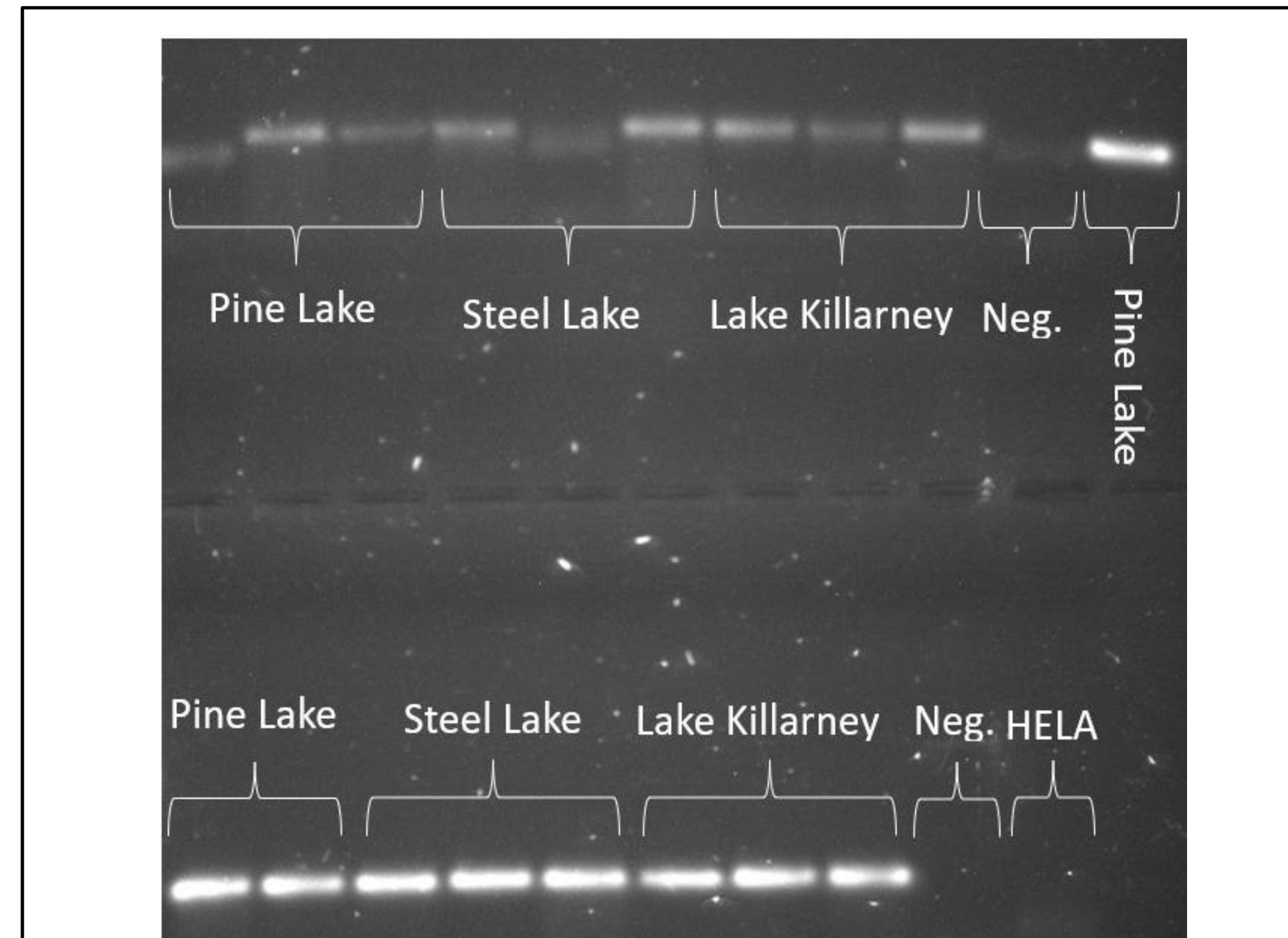
Genes were amplified and ran through the gel electrophoresis unit

Target gene was *hsp-70*

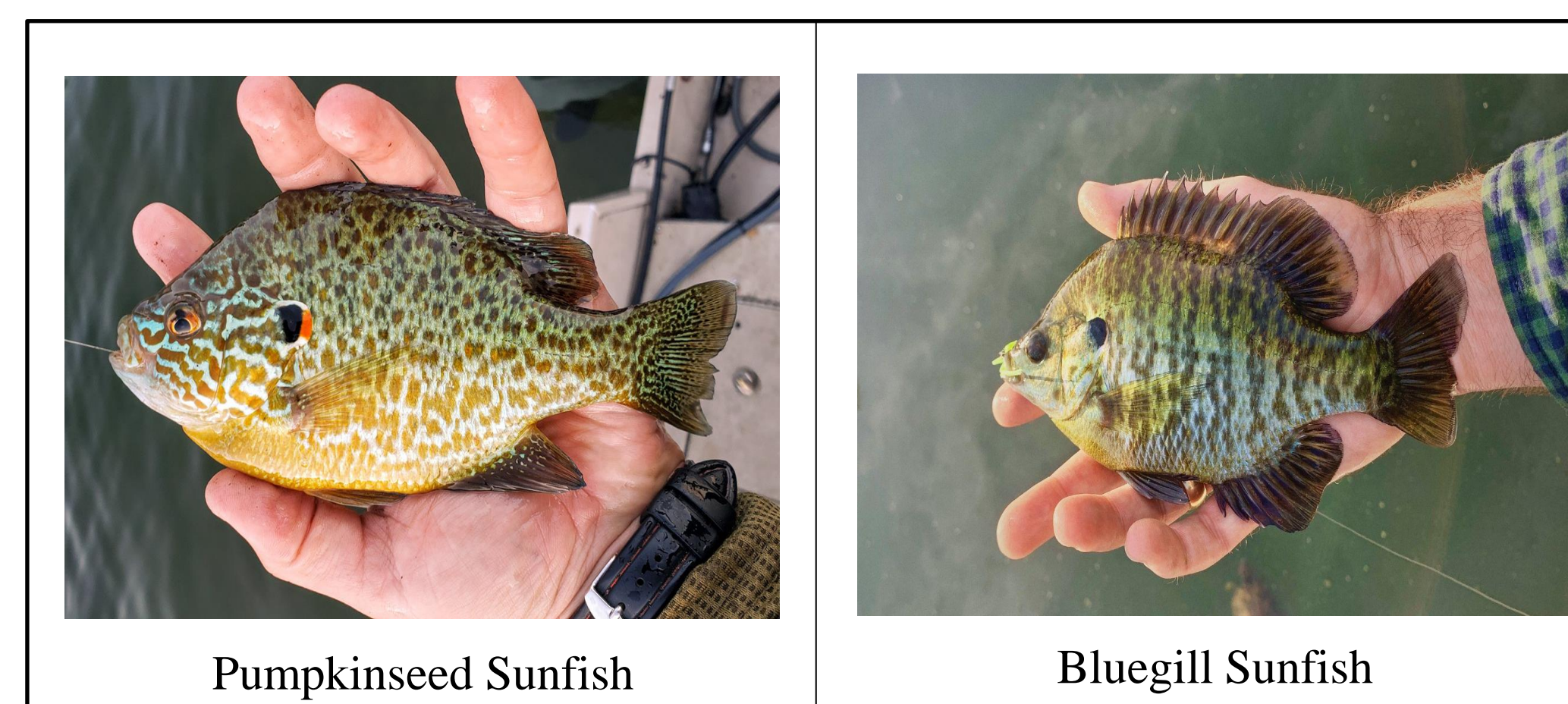
Housekeeping gene was *EF1-α*

Gels were analyzed under UV light and looked at the densitometry  
Used Image-J to find area of each band

Used ANOVA using the lakes as the single factor

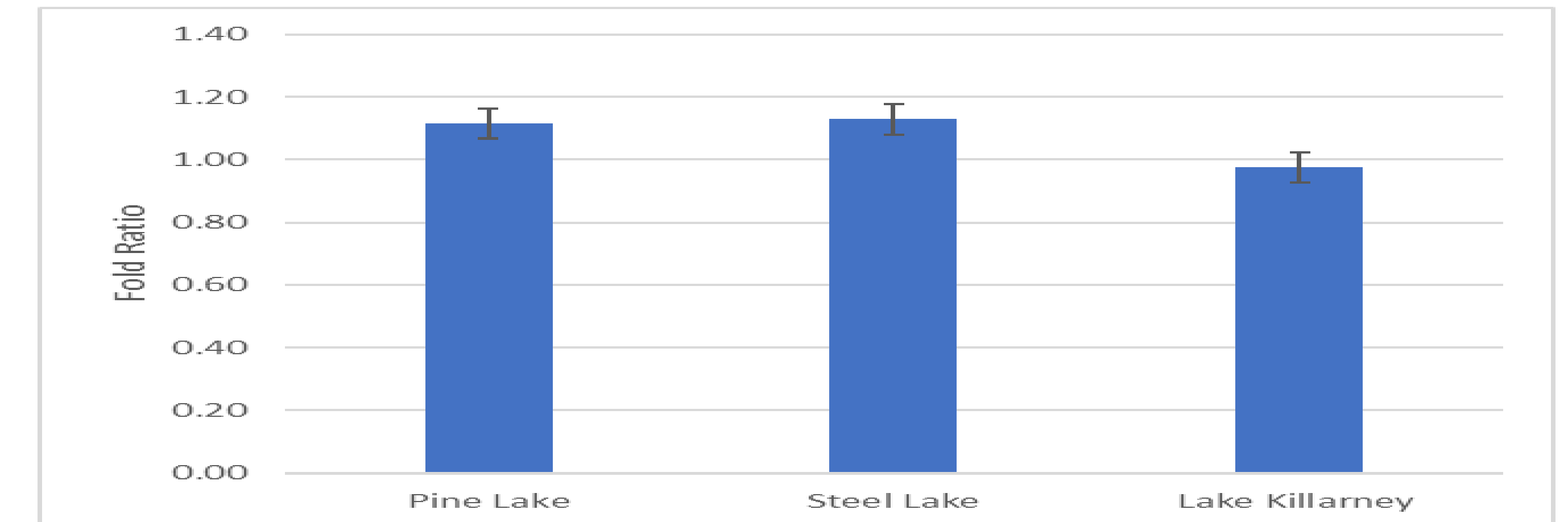


**Figure 1:** This image shows PCR amplicons of the fish collected from each of the lakes, Pine Lake, Steel Lake, and Lake Killarney. Both the target (*hsp-70*) and housekeeping (*EF1-α*) genes are shown. There was no significant difference in the *hsp-70* gene expression (Figures 1 and 2) across the three lakes. The first fish replica area from Pine Lake was not included in the calculations because the area was not able to be calculated accurately, as seen in the figure above the first replica sample was in the top row with the target gene samples so an accurate area was not able to be taken.



Pumpkinseed Sunfish

Bluegill Sunfish



**Figure 2:** This graph shows the fold ratio between the three lakes, Pine Lake, Steel Lake, and Lake Killarney. The fold ratio was calculated by dividing the areas between the target and housekeeping genes of each of three lakes to get the normalized area. And the normalized area of each area was divided by the average of the reference lake (Pine Lake) to find the fold ratio.

## RESULTS

There was no significant difference seen between the gene expression in the liver from any of the lakes in the Bluegill Sunfish and Pumpkinseed Sunfish with varying levels of arsenic contamination

P-value was 0.873, this is greater than 0.5 which also supports this claim

Error bars on the graph for fold ratio have significant overlap

One area for the housekeeping gene from Pine Lake was not included in the calculations to find these results

The results that came out did not support our hypothesis

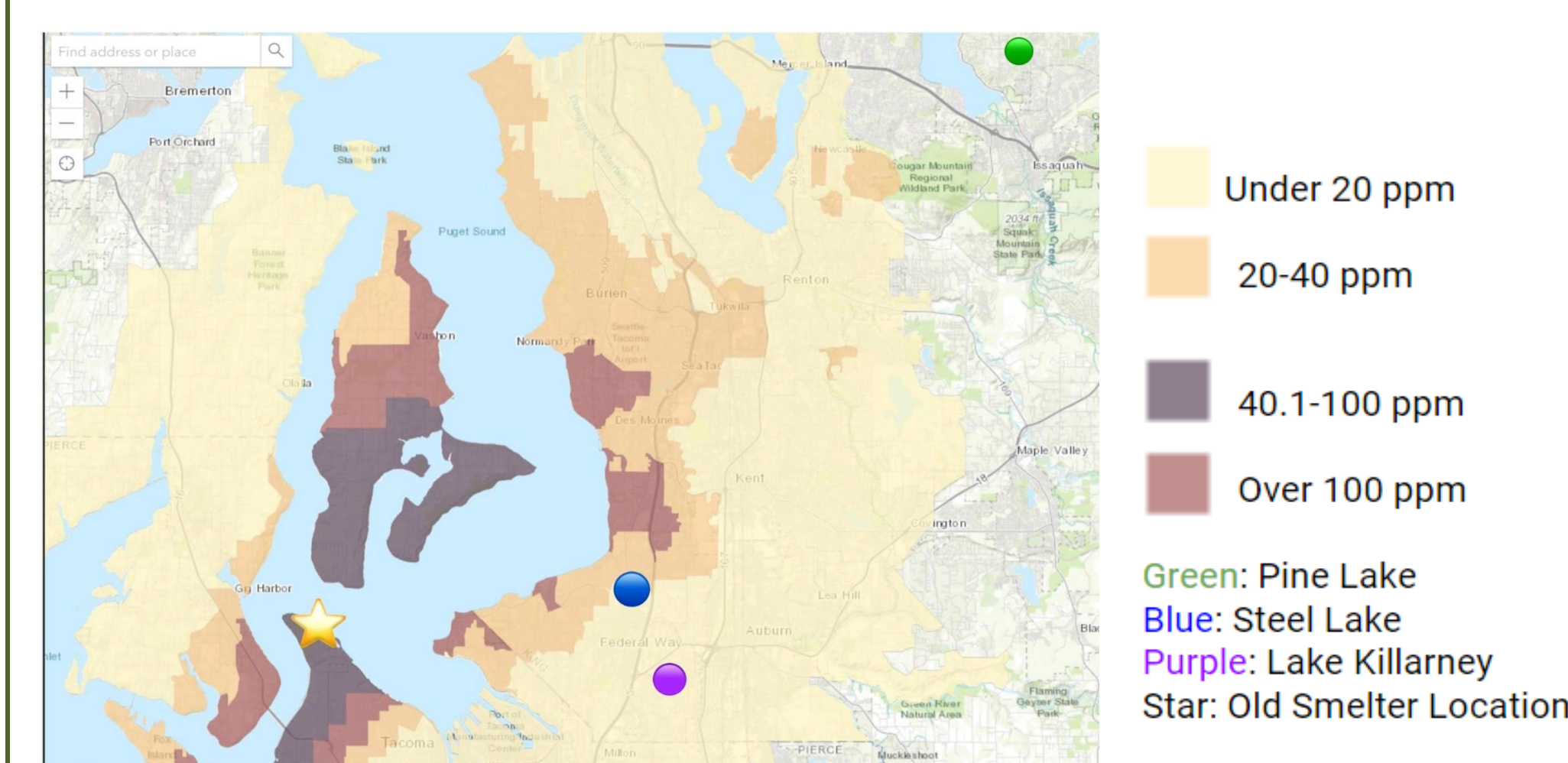
## DISCUSSION

We expected there to be a difference in the gene expression of the liver in the fish that were in the lake with higher levels of arsenic compared to the lakes with less or no arsenic contamination

There are still many other studies that can be performed to further study the effects of legacy contamination on fish, including analyzing biomarker responses in other tissues as well as other species.

Genes	Forward Primer	Reverse Primer
<i>Hsp-70</i>	AGA GAC TGA TTG GGA	TCC GTC TCC GAC CAC CTT
	GAAAGA TG	G
<i>EF1-α</i>	AAG CCT GGT ATG GTT	GCA TCT CAA CAG ACT
	GTG ACC TT	TGA CCT CAG T

Primer sequences acquired from (Dennis et. Al., 2015)



## REFERENCES

- "Arsenic." Washington State Department of Health, doh.wa.gov, doh.wa.gov/community-and-environment/contaminants/arsenic. Accessed 5 May 2022.
- Basu N, Todgham A., Ackerman P, Bibeau M., Nakano K, Schulte P, Iwama GK. 2002. Heat shock protein genes and their functional significance in fish. *Gene*. 295(2):173-183. doi:10.1016/S0378-1119(02)00687-X.
- Dennis CE, Kates DE, Noatch MR, Suski CD. 2015. Molecular responses of fishes to elevated carbon dioxide. *Comparative biochemistry and physiology Part A, Molecular & Integrative Physiology*. 187:224-231. doi:10.1016/j.cbpa.2014.05.015.
- GAWEL JIM, ENDRESEN ANTHONY, HULL ERIN, KEDIR AMINA, KING COREY, NEUMANN REBECCA, BARRETT PAM. Arsenic in lakes in the puget sound lowlands: Mobility and bioavailability 30 years after smelter closure. Washington State Lake Protection Association. 2016 Jun [accessed 2022 Jun 5]. <https://www.walpa.org/walpa-line/june-2016/arsenic-in-lakes-in-the-puget-sound-lowlands-mobility-and-bioavailability-30-years-after-smelter-closure/#:~:text=Our%20four%20current%20study%20lakes,shallow%2C%20well%2Dmixed%20lakes>.
- Infante C, Asensio E, Caffavate JP, Manchado M. 2008. Molecular characterization and expression analysis of five different elongation factor 1 alpha genes in the flatfish Senegalese sole (*Solea senegalensis* Kaup): differential gene expression and thyroid hormones dependence during metamorphosis. *BMC molecular biology*. 9(1):19-19. doi:10.1186/1471-2199-9-19.
- Wang Y, Guo B. 2019. Adaptation to extreme environments: a perspective from fish genomics. *Reviews in fish biology and fisheries*. 29(4):735-747. doi:10.1007/s11160-019-09577-9.

## ACKNOWLEDGEMENTS

The University of Washington Tacoma science department for making an awesome environment in which to research, work, and learn.

Dr. Jim Gawel and Washington Department of Fish and Wildlife

Dr. Alison Gardell for diligently directing the project.