

Adjusting the makeup of Glycan proteins to improve IgG BBB permeability

Fahad Kedir, Fernando A. Palomera Rubio, John M. Finke

Division of Sciences and Mathematics, Interdisciplinary Arts and Sciences, University of Washington Tacoma, Tacoma, WA, USA

INTRODUCTION

Alzheimer's disease (AD) affects around 10% of the world's population, and it is estimated that within the next 50 years, this diagnosis is expected to to triple. Alzheimer's disease is a neural disorder in which proteins are degrading neural cells and connections. The permeability of antibodies in the brain is very low. This makes it difficult to attack neural diseases in a conventional manner. IgG antibodies are a popular class of immunotherapy and they are used in clinical trials of AD. β -amyloid peptides are found in the brain of AD patients and these kinds of therapies are used to target and eliminate these peptides, as they are the cause of neural degradation. The only issue with this method is that IgG antibodies are not transported effectively in the brain. Dr. Finke in previous studies found that Fab α2,6-sialylated glycans on anti-amyloid IgG antibody 4G8 show a lower BBB efflux without a lower influx.

Blood Brain Barrier Studies of IgG Sialic acid

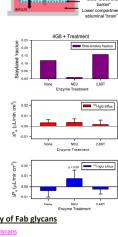
Previous in the Finke lab show that Fab α2,6-sialylated glycans on anti-amyloid IgG antibody 4G8 correlate with lower BBB efflux but not influx². IgG sialic acid may enable better IgG drug retention in the brain.

Treatment of 4G8 with neuraminidase

- 1. Removed all sialic acid (top).
- 2. Did not alter the influx rate (middle)
- 3. Reduced the efflux rate (lower)

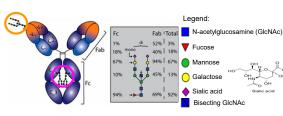
Treatment of 4G8 with \(\alpha 2.6\)-sialyltransferase

Did not significantly alter these parameters.



Antibodies have a diverse array of Fab glycans

Fab glycans All IgG have 2 Fc sialic acid cannot be detected in intact IgG. PNGase F will always cleave PNGase F cannot cleave unless IgG is "denatured".

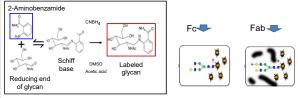


(S with HO or "g" indicates glycolated Sialic acid)

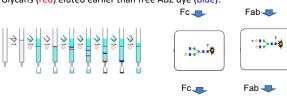
PROECT GOAL: Make a single sialylated form (G2FS1)

METHODS Glycan analysis - general process Analysis of Fab/Fc glycans 1. Glycan Release of Fc glycans (native conditions). Microcon 30kD ultrafiltration 2. Isolation of cleaved Fc glycans by 34 Microcon 30 kD filtration. 150 kD 2.5 kD PNGase F 3. PNGase F again on IgG+Fab glycans but with SDS, ß-mercaptoethanol, NP-40 at pH 8.6 (denaturing conditions). 4. Fc and Fab glycans were lyophilized and labeled via reductive

amination using 2-aminobenzamide and CNBH₄ reductant.



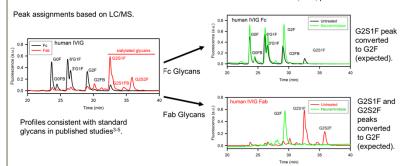
5. Labeled glycans were purified with size exclusion chromatography. Glycans (red) eluted earlier than free ABZ dye (blue).



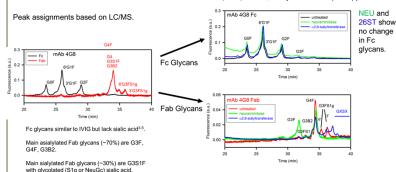
6. HPLC profile of glycans with polar glycan column. Glycans eluted between 20-40% ammonium formate pH 4.5 in acetonitrile. Measured with ABZ fluorescence.

HPLC Profiling of commercial 4G8 and Enzyme-Treated 4G8





Effects of neuraminidase (NEU) OR α 2,6 sialyltransferase (26ST) pre-treatment.



Results

We didn't register any peaks in our wavelenght graph, meaning there was not any or enough product to be analyzed. Control (above) with untreated 4G8 worked so outcome likely due to low levels of enzyme product after purification steps. These steps reduce our material so much that we get no results from them.

Possible solutions:

- 1. Single enzyme step (α 2,6-sialyltransferase + α -galactosidase together).
- 2. No purification steps (we'll figure this out if we see our product).

mer's Association. 2017; 3(4):325-373. Blochimica Et Blophysica Acta (BBA) - General Subjects munity." The Journal of Immunology, 2016; 196: 1435. a and its fragments Fab and Fc." Journal of Immunlogics General Subjects, 2017, 1861; 2228-2239., doi:10.1016/j.bbagen.2017.06.008. nicel Methods 2012: 382:167-176

National Institutes of Health / NIA R03 AG050184 (JMF, WAB)

M.J. Murdock Charitable Trust (JMF, ED)