



# Effect of *Porphyromonas gingivalis* outer membrane vesicles on biofilm dispersal through modification of Fimbria and Lipid A phosphorylation



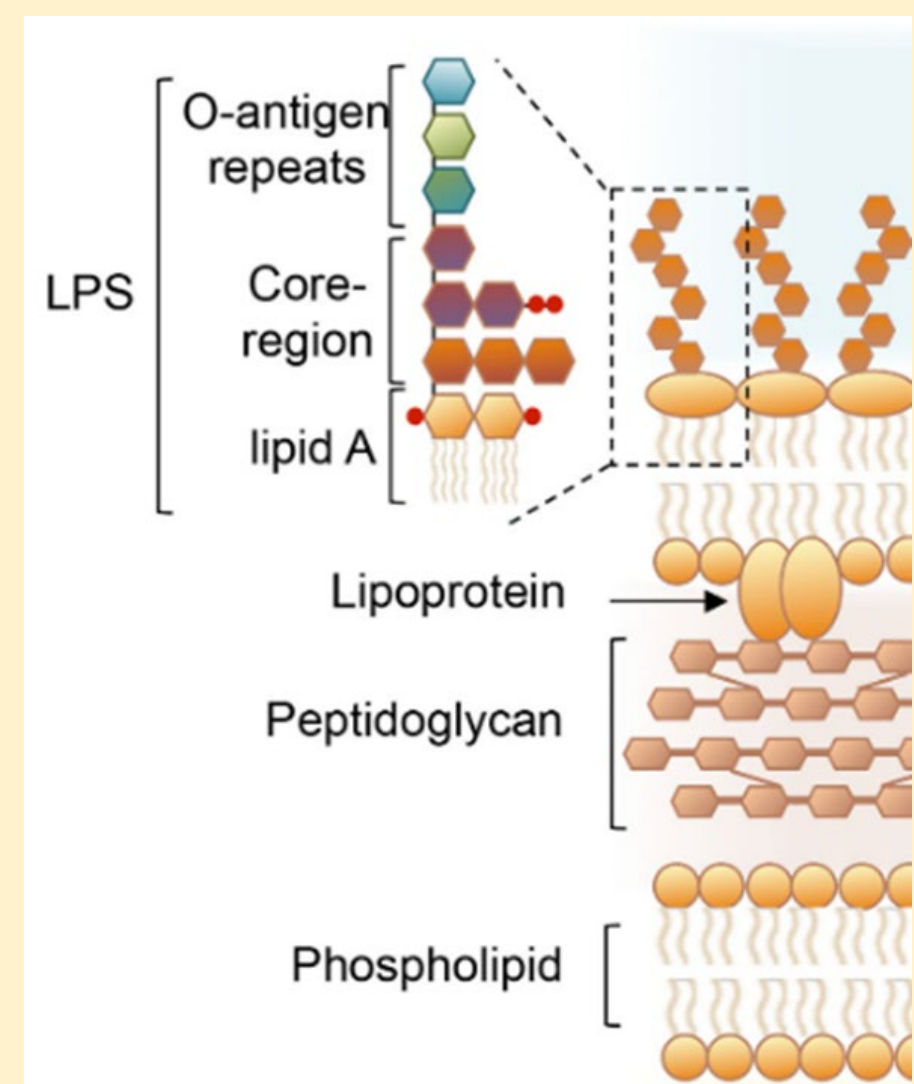
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## Introduction

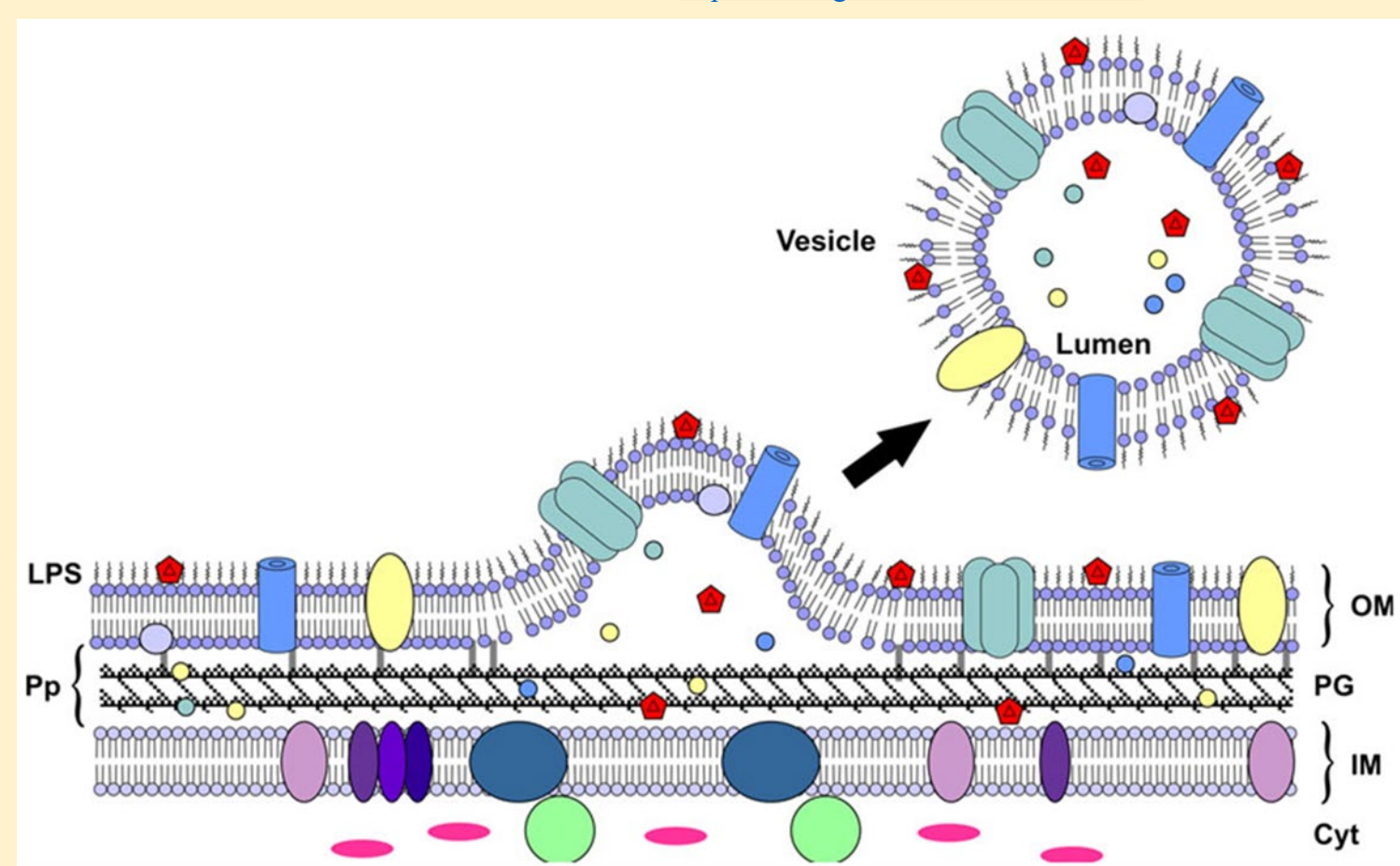
- *Porphyromonas gingivalis* is an anaerobic gram-negative bacteria that resides in the gum layer of the oral cavity and correlated with the progression of periodontal disease.
- The lipopolysaccharide is a conserved tripartite structure across gram-negative bacteria and serves in membrane protection and stabilization.
- Gingipains – *P. gingivalis* virulence factors that aid in bacterial establishment through the dispersal of non-self bacteria.
- Hypothesis #1: If Lipid A is modified via phosphorylation, then it will affect the outer membrane vesicle's ability to disperse biofilm.
- Hypothesis #2: If fimbria were knocked out from *P. gingivalis*, then that will reduce outer membrane vesicle-mediated biofilm dispersal.

## Background

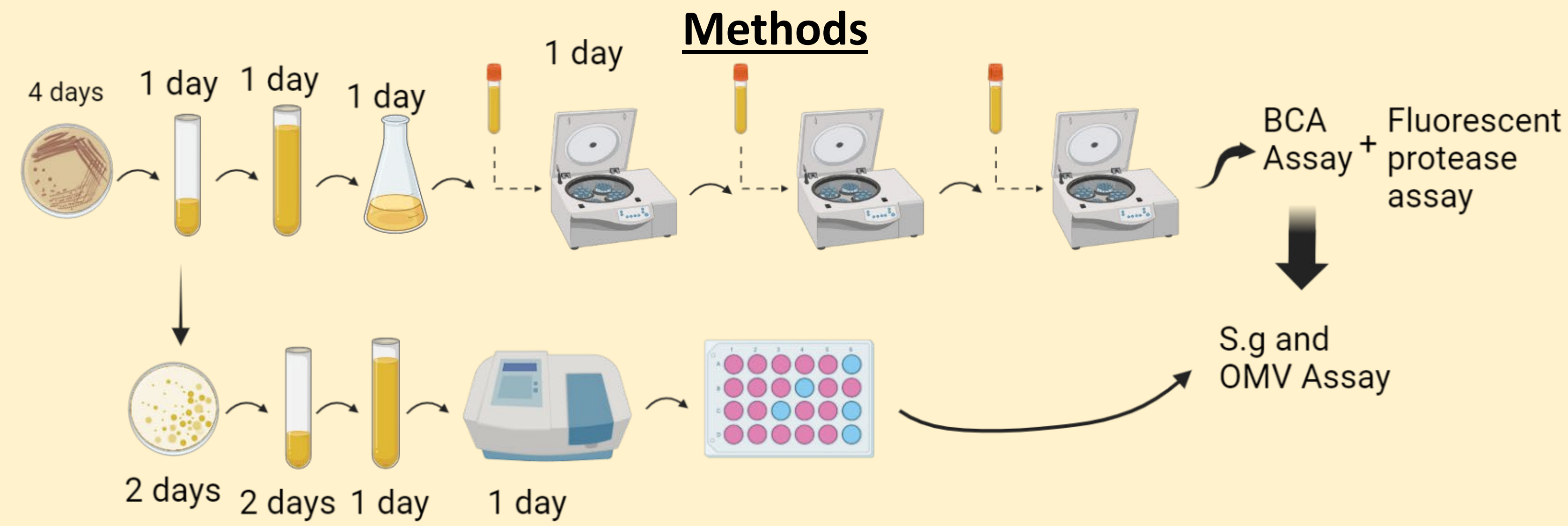
- What is a lipopolysaccharide?
- What is an outer membrane vesicle?
- How do they tie together?



Rita F. Maldonado, Isabel Sá-Correia, Miguel A. Valvano, Lipopolysaccharide modification in Gram-negative bacteria during chronic infection. *FEMS Microbiology Reviews*, Volume 40, Issue 4, July 2016, Pages 480–493, <https://doi.org/10.1093/femsre/fuv007>



Gui, M.J., Dashper, S.G., Slakeski, N., Chen, Y., & Reynolds, E.C. (2016). Spheres of influence: *Porphyromonas gingivalis* outer membrane vesicles. *Molecular oral microbiology*, 31 5, 365-78.



## Results

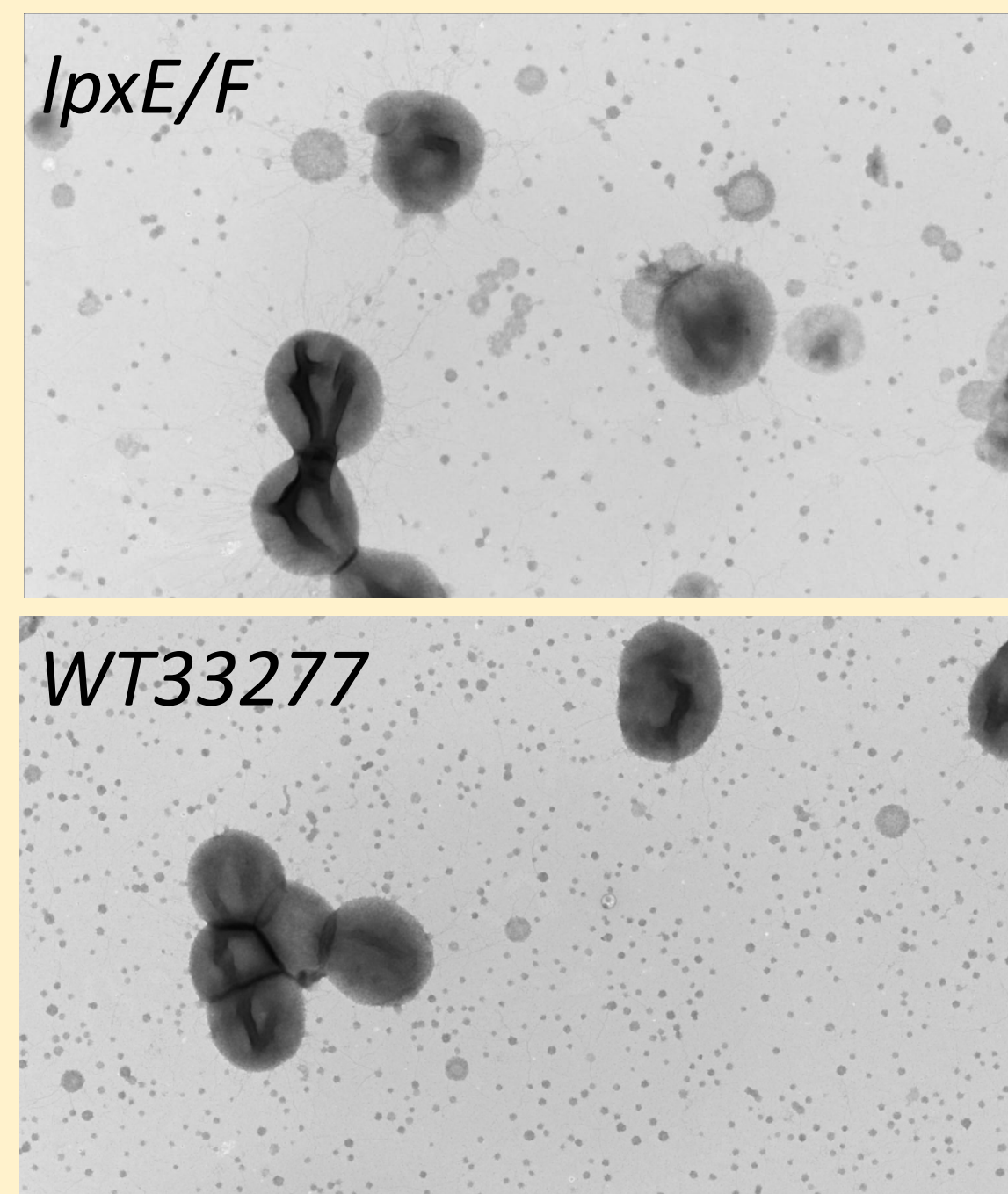


Figure 1: TEM pictures of *lpxE/F* & WT33277. *lpxE/F* shows a depletion of OMVs when compared to WT33277.

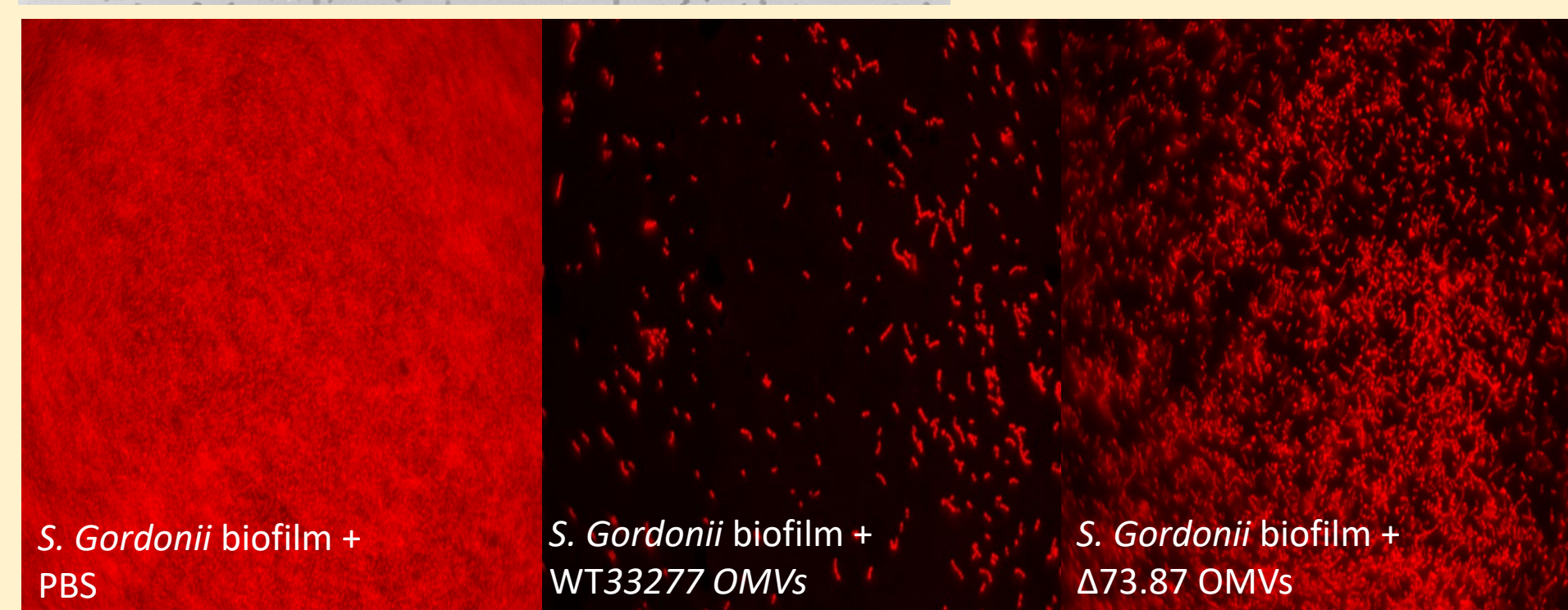


Figure 2: *Streptococcus gordonii* & *Porphyromonas gingivalis* OMV biofilm. The left panel shows the control. While the middle panel with WT33277 OMVs depicts biofilm dispersal. While the panel on the right depicts less dispersal.

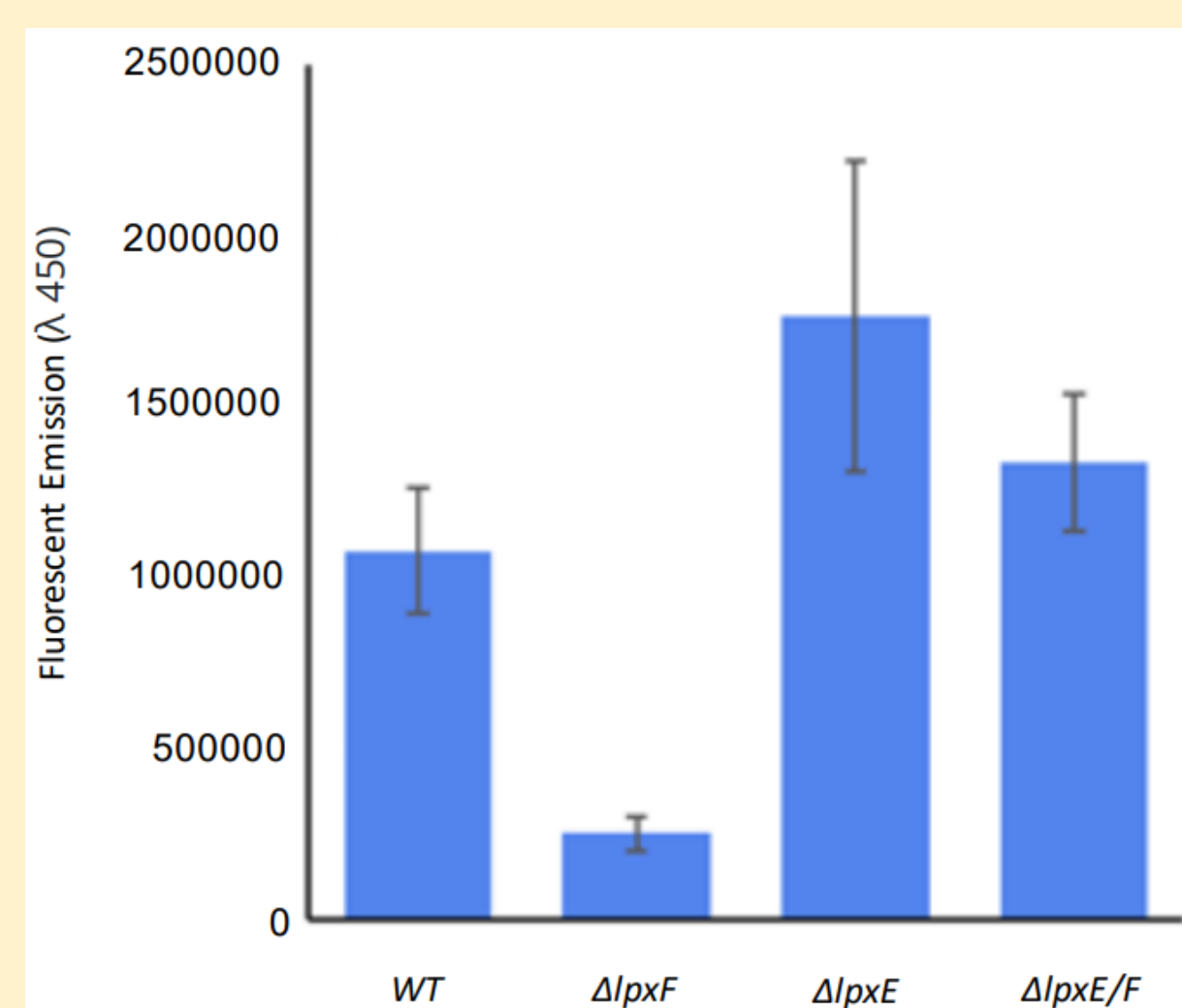


Figure 3: DPH outer membrane vesicle quantification. WT33277 & *lpxE/F* show a similar production of OMVs.

WT1 (mP)		lpxE/F (mP)	
1:2	-60.864004245	1:4	-54.940208247
1:4	-56.525996063	1:16	-48.352352552
1:8	-53.577554051	1:32	Uncounted
WT2 (mP)		lpxE/F (ΔmP)	
1:4	-56.124665597	1:2	-0.03130594744
1:16	-46.027963726	1:2	-0.01322052007
1:32	-36.717798512	1:5	-0.0106445741
ΔGingi (mP)		1:10	-0.00369021499
1:4	691.893240941		
1:16	442.360075812		
1:32	7.223609041		
WT1 (ΔmP)		lpxE/F (ΔmP)	
WT	-0.02477633775	lpxE/F	-0.03130594744
1:2	-0.01572600639	1:2	-0.01322052007
1:5	-0.00527758622	1:5	-0.0106445741
1:10	-0.01503046341	1:10	-0.00369021499

Figure 4: Protease assay of outer membrane vesicles. WT1 shows a dilution series of 1:2 – 1:8. WT2 & *lpxE/F* show a dilution series of 1:4 – 1:32. Gingi is a control. Repeat WT1 & *lpxE/F* shows a dilution series of 1:2 – 1:10. Dilution 1:10 shows a 5x difference in fluorescent polarization.

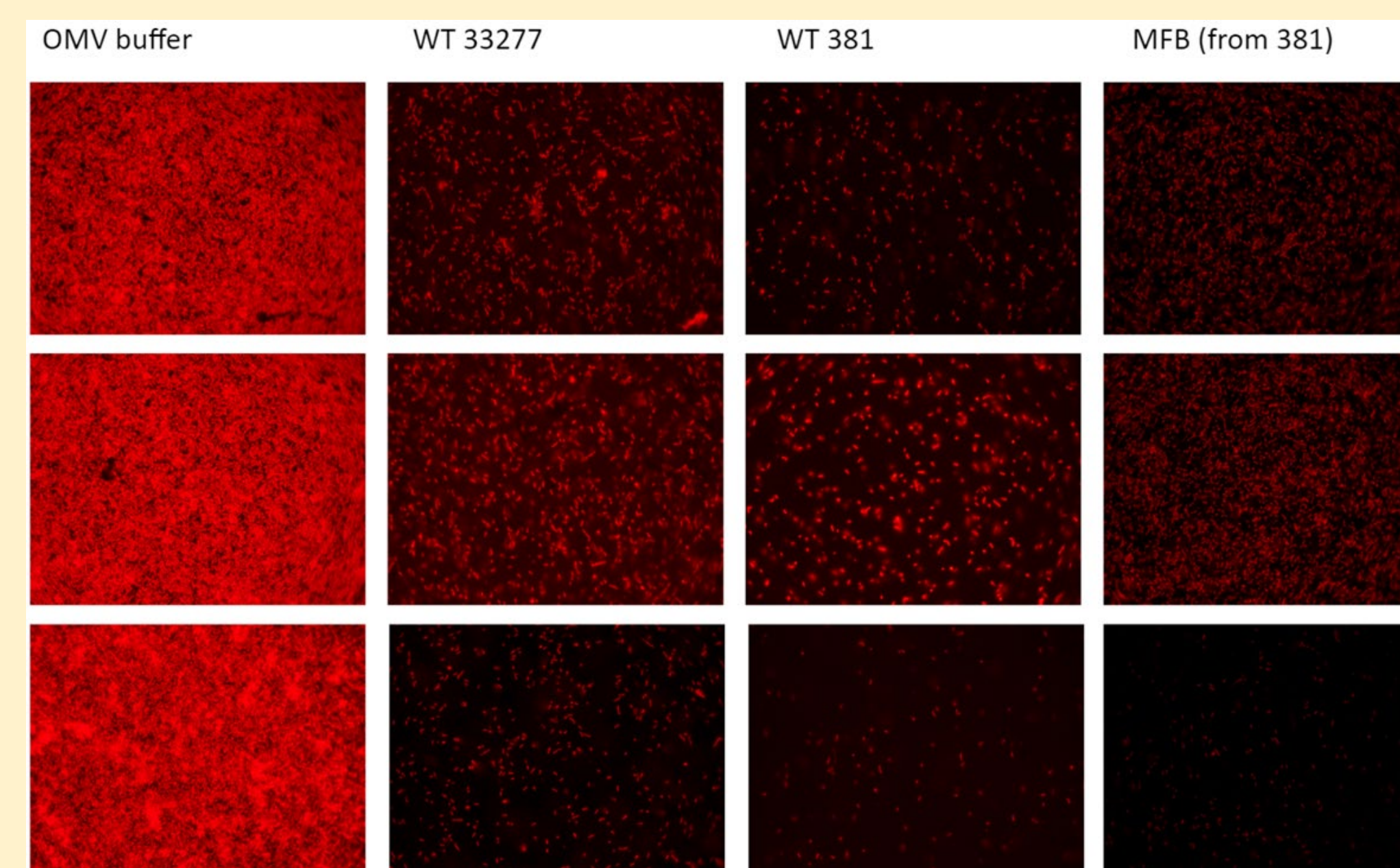


Figure 5: *P. gingivalis* introduction to *S. gordonii* biofilm. Column 1 shows *S.g* biofilm with 250 ul buffer as a negative control. Column 2 shows 15.09 ul of strain 33277 plus 235 ul buffer in *S.g* biofilm. Column 3 shows 150 ul of strain 381 plus 227 ul of buffer in *S.g* biofilm. Column 4 shows 23 ul of strain MFB plus 145 ul of buffer in *S.g* biofilm.

## Conclusions

- Hypothesis #1: Preliminary data of the protease activity suggest that activity decreases with an increase in protein dilution. However, due to inconsistent data using the Fluorescent Protease Assay Kit, we concluded it was not an optimal measure to determine protein activity with OMVs. Future research should be directed toward optimizing an assay that adequately measures protease activity in a consistent manner.
- Hypothesis #2: If fimbriae deletion occurred then there should be little-to-no biofilm dispersal of *Strep*. This hypothesis proved to be inaccurate since the MFB strain was able to effectively disperse *strep* biofilm. Note: there was a miscalculation during the S.g + OMV assay as fewer MFB strain quantity was added into the biofilm. Since dispersal was concluded at a much smaller volume, and in conjunction with literature review; fimbria deletion consideration was no longer investigated.
- *P. gingivalis* morphology characterization and outer membrane vesicle production had been consistent up until Winter 2023. Upon receiving new bacterial stocks from UW Seattle, *lpxE/F* is now producing much fewer OMVs than previously characterized while WT33277 is producing longer fimbria units.
- As of late, the lab has been focusing on characterizing the morphology of different *P. gingivalis* strains and will investigate genomic sequencing.
- Understanding *P. gingivalis* OMVs and their role in biofilm dispersal, future research could be directed toward OMV biogenesis, cessation, and progression of periodontal disease.

## Acknowledgments

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